

In-vitro Anthelmintic Efficacy of the 80% Hydro-alcohol Extract of *Myrsine africana* (kechemo) Leaf on Hookworm Larvae

Basha H*, Debella A, Hailu A, Mequanente S, Mersa A, Ashebir R, Degu S, Mesay GMT, Tadele A, Biruktawit G, Sahle A and Netsanet F

Ethiopian Public Health Institute, Traditional and Modern Medicine Research Directorate, Ethiopia

*Corresponding author: Basha H, Ethiopian Public Health Institute, Biomedical and Clinical Research Team, Traditional and Modern Medicine Research Directorate, P.O.Box 1242, Addis Ababa, Ethiopia, Tel: +251 0910621573, E-mail: hirut_basha@yahoo.com

Citation: Basha H, Debella A, Hailu A, Mequanente S, Mersa A, Ashebir R, et al. (2018) *In-vitro* Anthelmintic Efficacy of the 80% Hydro-alcohol Extract of *Myrsine africana* (kechemo) Leaf on Hookworm Larvae. J Public Health Dis Prev 1: 103

Article history: Received: 15 August 2018, Accepted: 10 October 2018, Published: 12 October 2018

Abstract

Background: Helminthic infections are among the most common infections in human, affecting a large proportion of population all over the world. Among the most important helminthes found in Ethiopia are *Ascaris*, Hookworms, Schistosomes, Strongyloides, *Trichuris* and *Taenia saginata*. Hookworms affect a large number of people in Ethiopia. Although the prevalence of hookworm and other parasitic infections are high in Ethiopia there are not enough medical services and modern medicine to treat the parasitic infection. The majority people in Ethiopia rely on traditional medicine to treat worm infestation.

Objective: The aim of this study was to evaluate *in vitro* larvicidal activity of the 80% Hydro-alcohol extract of *Myrsine africana* and determine the phyto constituents contained in the plants through the general phytochemical investigation.

Methods: A phytochemical screening for *M. africana* leaf extract was done using detecting reagents for the presence of alkaloids, glycosides, steroids, saponins, flavonoids, terpenoids, and tannins among others. And *In vitro* larvicidal effect on the third stage larvae of hook worm was carried out using a 96-well microtiter plate assay. In- vivo acute toxicity study was also done on Swiss albino female mice.

Result: The preliminary phytochemical screening analysis revealed the presence of alkaloids, tannins, polyphenols, steroids, saponin and glycosides on the 80% Hydro alcoholic leaf extract of *M. africana*. The result also indicated that 80% Hydro alcoholic extracts of *M. africans* exhibited larvicidal activity against hookworm larva. The LC₅₀ was 217.77 microgram per milliliter.

Conclusion: *Myrsine africana* appear to possess promising anthelmintic activity that may support the usage of the plant by local traditional healers to treat hook worm infection.

Keywords: Anti-helmentic; Hookworm; *Myrsine africana*

Abbreviations or Acronyms: EPHI: Ethiopian Public Health Institute; TMMRD: Traditional and Modern Medicine Research Directorate; SNNPR: Southern Nations Nationalities and Peoples Region of Ethiopia; OPD: Out Patient Department; DMSO: Dimethyl Sulfoxide; L3: Filariform Larva; RPM: Revolution per Minute; µg: microgram; ml: milliliter; LC: Lethal Concentration; IRB: Institutional Review Board; mg: milligram

Introduction

Helminthic infections are among the most common infections in human, affecting a large proportion of population all over the world. In developing countries they pose a large threat to public health and contribute for malnutrition, anaemia, eosinophilia and pneumonia and cause malabsorption, diarrhoea, and other states of poor health, particularly in infants and school-age children [1-4]. Infections due to intestinal parasites are common throughout the tropics, posing serious public health problems in developing countries which may be attributed largely to socio-economic status, poor sanitation, inadequate medical care and absence of safe drinking water supplies [5]. Globally, 3.5 billion people are affected by intestinal parasites [6]. Approximately 300 million people suffer severe morbidity associated with helminthic parasites and half of which are school-going children [7]. Helminthic diseases also pose a major health hazard to millions of livestock and cause significant economic losses in domestic and farm animals [4].

A study reported that the prevalence of intestinal parasitic infections in developing countries is almost twice those of developed countries [8]. With their high prevalence in Ethiopia, Parasitic helminthic infections cause serious public health problems [9]. More than half million visits of OPD in the country were due to intestinal parasitic infections [6]. *Ascaris*, Hookworms, Schistosoma, Strongyloides, Trichuris and *Taenia saginata* are among the most important helminthes found in Ethiopia. Hookworms affect a large number of people in our country [10].

The intensive use of anthelmintics for the control of helminthic infections has resulted in the development of anthelmintic resistance, which has become a major practical problem in many countries [11]. The high prevalence of intestinal parasitic infections, the lowering efficacy of current chemotherapies due to the appearance of resistant species inadequate medical services, unaffordability and inaccessibility of modern anthelmintics particularly to the rural population have awakened the interest of medicinal plants as an alternative medicine for control of these parasites [11-15]. Thus, an urgent need for the investigation and development of effective, safer, inexpensive anti-helminthic drugs from medicinal plants is very crucial [16-17]. Around 80% of the Ethiopian population relies on plant derived remedies to satisfy their health care needs [18]. Traditional herbal remedies such as the seeds of *Embelia schimperi* (Enkoko in Amharic), *Maesa lanceolata* (Kelewa in Amharic), *Hagenia abyssinica*, the bulb of oxalis anthelmintica (Michamcho in Amharic), the berries of *Myrsine africana* (Kechemo in Amharic) and the roots of *Punica granatum* (roman in Amharic) etc have been used in treating tapeworm infestations for ages [6].

Myrsine africana is selected for the current study. This plant belongs to the Myrsinaceae family that has been used traditionally by many of the country's ethnic groups for treating various ailments such as tape worm [6,19]. Myrsine family, is a rather large family from the order Ericales. It is composed of more than 1000 species in over 30 genera worldwide. It is a widespread family belonging to temperate to tropical climates extending north to Europe, Siberia, Japan, Mexico, and Florida and south to New Zealand, South America, and South Africa [16]. *Myrsine africana* found in different parts of Ethiopia: Tigray, Gondar, Gojam, Wello, Arsi, Shewa, Welega, Kefa, Sidamo, Bale and Hararge [20]. People mix *Myrsine africana* dry leave powder with honey and take it orally to treat round worm and tapeworm.

Thus, the present study is intended to evaluate the *in vitro* larvicidal activity of the 80% Hydro alcoholic extract of *Myrsine africana* on hookworm larvae and identify the phytoconstituents contained in it.

Materials and Methods

Drugs and Chemicals:

Albendazole are used as a standard drug (as positive control) during the experimental protocol and 1% DMSO was used as negative control. All the chemicals and reagents used are laboratory and analytical grade.

Plant material collection:

The fresh leaves of *Myrsine africana* were collected from Jimma zone located 352.4km from Addis Ababa. The plant material was identified and authenticated by a taxonomist at the Traditional and Modern Medicine Research Directorate (TMMRD) of the Ethiopian Public Health Institute (EPHI). A voucher specimen (No-2177) was deposited for future reference.

Plant parts preparation and extraction:

The plant leaves were washed with running tap water, dried in open air and ground to powder for extraction. Two hundred grams of *Myrsine africana* powdered leaves were extracted with 80% Methanol using an electrical shaker for three consecutive days. The extract was filtered using what man number 1 filter paper and concentrated by evaporation using rotary vaporizers under reduced pressure at a temperature of 40-45 °C. The filtrate obtained was then dried by steam bath at 40 °C and weighed and, finally kept in a refrigerator at 4 °C for experimental usage.

Qualitative Phytochemical tests of plant extract:

General Phytochemical screening tests were carried out for the *M. africana* 80% crude hydro alcoholic leaf extract following procedures adopted from Harborne [21-22]. The extract was tested for the presence of alkaloids, glycosides, steroids, saponins, flavonoids, terpenoids, and tannins among others and identified by characteristic colour changes using standard procedures.

Test for Saponins: Crude extract was mixed with distilled water and heated for 5 minute, then filtered. The filtered solution was shaken vigorously for 2 minute. Foam that doesn't disappear was taken as indication of the presence of saponin in the extract.

Test for Polyphenol: Test solution was treated with few drops of a mixture of 1 ml each of 1% FeCl₃ (ferric chloride) and 1% K₃Fe (CN)₆ were added to 2 ml of the aqueous solution of the extracts. Formation of green or blue color was taken as an indication of the presence of polyphenols.

Test for Glycosides: Crude extract was macerated in distilled water to form a thick mass. A piece of filtered paper was dipped in 1% Ag picric acid and 10% Na₂CO₃ and then suspended above the thick mass. Formation of brick red colour on the filter paper indicates the presence of cynogeneic glycosides.

Test for Alkaloids: Crude extract was mixed with 10ml of 1% HCl (hydrochloric acid) and heated for 30 minutes. It was cooled and filtered. To 1ml of the filtrate, 5 drops each of Mayer's and Dragendorff's reagents were added. Formation of white, yellow orange and white precipitate respectively indicated the presence of alkaloids in the extract

Test for Tannin: 2ml of ethanol extract was mixed with a few crystals of sodium nitrate. 3 drops of 0.1N HCl was added to extract solution. The formation of brown colour indicates the presence of tannin.

Test for Steroids: 1.5% vaniline spray: The spray reagent is prepared by mixing equal volume of ethanol and sulphuric acid. Spray the TLC plate with reagent followed by heating in an oven at 90-100 °C for 2-3 minutes. Brown spot indicates the presence of steroid.

Standard solution: Albendazole (200, 100, 50, 25 and 12.5 µg.mL⁻¹) prepared from 20,000 µg.mL⁻¹ of previously prepared stock solution) was administered as standard solution.

Test solution: Different concentrations (1000, 500, 250, 125 and 62.5 µg.mL⁻¹) of methanol leaves extracts of *M. africana* were prepared from 100,000 µg/ml of previously prepared stock solution. All the extracts and the standard drug solution were freshly prepared before starting the experiments.

Parasite acquisition: Hookworm parasites used in this study were obtained from a source population of 200 patients that visited Cheha Health Center Laboratory, Southern Nations Nationalities and Peoples Region of Ethiopia (SNNPR) located 180 km away from Addis Ababa. The site was chosen because it is known to be endemic for hookworm disease and has favorable conditions. Diagnosis of the parasite was undertaken by standard microscopic techniques. A faecal sample that tested positive for both *Strongyloides stercoralis* and hookworm was excluded from the study. The diagnosis of the parasite was cross checked by two Laboratory technologists for confirmation.

Larva preparation: Harada-Mori technique was used to hatch the hook worm larva as follows; 1 gm of faeces was placed on Whatman filter paper No.1 and placed in a 15-ml Centrifuge tube containing 4 ml of distilled water was added to the bottom of the tube. To avoid fungal contamination Nystatin, an anti-fungal agent 10mg/ml was prepared and three drops of this solution was added to each tube [23]. The Tubes were kept for approximately 10 days at room temperature. The larva examination was commenced on the fifth day. To obtain enough number of larvae, the fluid was concentrated by low speed centrifugation (2,000 rpm for 10 minutes). The numbers of larvae present was ascertained by counting an aliquot. The volume was adjusted to achieve a concentration of one larva per microliter.

In vitro larvicidal Assay: The assay was carried out using a 96-well microtiter plate following methodology described by Gill [24]. Stock solutions of 80% methanolic crude extract of *M. africana* and albendazole were prepared at 100,000 and 20,000 microgram/ml, respectively, in 1% DMSO and were serially diluted by two-fold to produce a series of dilutions. Aliquots (2µl) were added to 2% molten agar in a total volume of 200µl in individual wells of a 96-well micro titer plate. The final drug concentrations in the assay plates consisted of two-fold serial dilutions starting at 1000, 500, 250, 125 and 62.5 µg.mL⁻¹ for the plant and 200, 100, 50, 25 and 12.5 µg.mL⁻¹ for the standard drug Albendazole. 1% DMSO was used as a negative control. Approximately 30 larvae (in 30µl of distilled water) were added to each well, and the plates were maintained at a constant temperature of 25°C for 2 days. Then the larvae were stimulated to move by addition of 40µl of water warmed to 50 °C to each well; Individual larvae moving with a smooth sinusoidal motion were considered to be unaffected by the test substance and counted under a microscope. All assays were performed using triplicate assay wells at each drug concentration.

In vivo acute toxicity study: Swiss albino female mice with a weight range between 20 and 25 g were divided into two groups of six test animals each. Four hours prior to the experiment, the animals were deprived of food while water was given *ad libitum*. Mice in the negative control group received saline while the experimental group received a limit dose of 2,000 mg/kg of the 80% methanolic crude extract of *M. africana* orally. All animals were then strictly observed continuously for 24 hours, with due attention paid to the first 4 hours after treatment. Any overt signs of morbidity and mortality were recorded. The animals were kept for continual observation for up to 14 days thereafter [25].

Data analysis:

The dose-response data was analyzed using non-linear regression model (sigmoidal dose-response, GraphPad Prism; Graph Pad Software, Inc., San Diego, CA). Drug sensitivity data was expressed as lethal concentration (LC₅₀) values (with 95% confidence intervals). This is defined as the concentration of a drug required to kill 50% of the motile worms observed in wells of the micro titer plate.

Ethical Consideration:

Ethical approval was obtained from the ethics committee and Institutional Review Board (IRB), Faculty of Medicine, Addis Ababa University. All the study participants of the study gave informed consent before stool specimen was collected. Those with confirmed hookworm infection were treated free of charge.

Results

In vivo acute toxicity study:

The experimental animals showed no clinical signs of toxicity and overt behavioral changes at the oral limit dose of 2,000 mg/kg the crude 80% Hydro alcoholic leaf extract of *M. africana*. Likewise, no mortality was recorded during the observation period of 14 days. Thus, the lethal dose (LD₅₀) of the plant extract was beyond 2000mg/kg of body weight.

Phytochemical screening:

The percent yield of the extracts of *Myrsine Africana* was determined to be 8.4% (w/w). Results of preliminary phytochemical screening analysis revealed the presence of alkaloids, tannins, polyphenols, steroids, saponin and glycosides, from the 80% methanolic extracts of *M. africans*. The results are presented in Table 1.

Class of compound	<i>M. africana</i>
Saponins	(+)
Polyphenol	(+)
Steroids	(+)
Flavonoids	(-)
Alkaloids	(+)
Glycosides	(+)
Tannins	(+)

(+) = present; (-) = absent

Table 1: Result of qualitative phytochemical analysis

In vitro larvicidal assay

The 80% Hydro-alcohol extracts *M. africans* exhibited larvacidal activity against hookworm.

	LogLC ₅₀		LC ₅₀ (µg.mL-1)	
	Best fit	Std error	Best fit	95%CI
<i>M. africana</i>	2.338	0.03386	217.77	185.1 to 255.9
Albendazole	1.636	0.03318	43.25	36.66 to 51.03

Table 2: LC₅₀ of the crude extract and albendazole

The concentration required to produce 50% larval inhibition (LC₅₀) of the plant extract and the standard drug, albendazole were 217.77 and 43.25 µg.mL⁻¹ respectively (Table 2).

Concentration (µg.mL-1)	Albendazole	
	% Lethal	95% CI
12.5	22.2	13-31.5
25	28.5	17.8-38.9
50	44.4	35.2-53.7
100	66.7	57.4-76
200	91.5	86-96.7

Table 3: Percent mortality of larvae at different concentration of standard drug albendazole

The standard drug albendazole inhibit larvae activity of the infective stage (L3) of hookworms. As the concentration of standard drug increases, the percent of hook worm larva mortality also increases (Table 3).

Concentration (µg.mL-1)	<i>M. africana</i>	
	% Lethal	95% CI
62.5	22.2	13-31.5
125	26	20.4-38.8
250	44.4	35.2-53.7
500	81.5	72.2-90.7
1000	98.8	93.3-104

Table 4: Percent mortality of larvae at different concentration of hydroalcoholic extract of *M. africana*

The crude extract of *M. africana* significantly (P<0.05) inhibit larvae activity of the infective stage (L3) of hookworms. Observation of the dose dependent effect of the extract also indicates that as the concentration of *M. africana* increases, the percent of hook worm larva mortality also increases (Table 4).

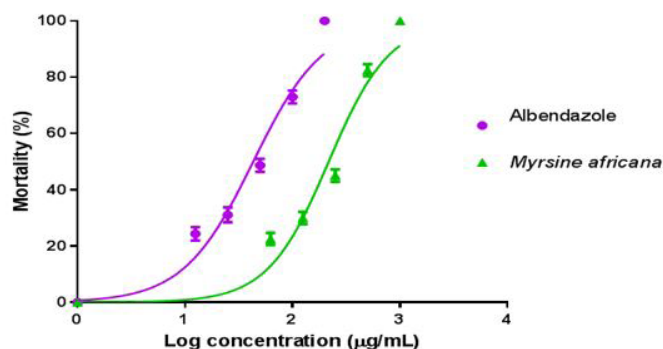


Figure 1: Larval mortality (%) vs. the base10 logarithm of the concentrations (µg/ml) the standard drug, Albendazole and the test solution of *Myrsine africana*

The result also indicated that 80% Hydro-alcohol extracts of *M. africana* exhibited anthelmintic activity in dose-dependent manner (Figure 1).

The mean percentage mortality was plotted against the logarithm of concentrations and the concentration killing 50% of the larvae (LC_{50}) were determined from the graph using Microsoft Excel 2013 computer software.

Discussion

Traditional medicines hold a great promise as source of easily available effective anthelmintic agents to the people, particularly in developing countries. The origin of many effective drugs has been found in the traditional medicines practices and in view of this it is important to undertake studies pertaining to screening of the traditional medicinal plants for their proclaimed anthelmintic efficacy. This study tries to evaluate *in vitro* helminthocidal activity of *M. africana* [26]. The use of *in vitro* assay to evaluate antihelminthic activities of plants and plant extracts has two main advantages: lower cost compared to *in vivo* experiments and it can be conducted without interfering the physiological function of the host [27].

The acute toxicity test done on mice at limit dose of 2000 mg/kg indicated the safety of the 80% methanol extract of the plant as no overt signs of morbidity and mortality were recorded during the experimental period. In this experiment the result revealed that hydro alcoholic extract of *Myrsine africana* exhibited larvicidal effects on the hookworm larva. The LC_{50} concentration larval inhibition of the plant extract and the standard drug, albendazole were 217.77 and 43.25 µg.mL⁻¹ respectively. The concentration of extract seems a bit higher compared to standard drug, this is because extracts were crude that constituent many chemical activity. There is a need for further study to identify which specific chemical constituent has anthelmintic activity and to understand the mechanism of action.

The larval development hasn't been affected in the absence of the plant extracts and albendazole. Although there are no studies conducted to evaluate *Myrsine africana* effects on hookworm larva, several studies indicated that *Myrsine africana* has anthelmintic activity [28-32]. According to a study conducted by [33]. *Taenia saginata* was expelled from humans after dosing with aqueous or ethanolic extracts of *M. africana* fruits. An *In vitro* study by demonstrated that alcoholic extracts of *M. africana* fruits were highly efficacious against the nematode *Bunostomum trigonocephalum* and the cestode *Taenia solium* [31].

Previous studies have declared the importance of some secondary metabolites such as alkaloids, glycosides, terpenoids, tannins and flavonoids for showing anthelmintic activity of medicinal plants [4,33-34]. The presence of flavonoids, glycosides, saponins, alkaloids, tannins etc in the hydro- methanolic extract of *M. Africana* are were responsible phytochemical constituents for demonstrating anthelmintic activity of the methanolic extract of the plants as these groups of metabolites are detected. As a result, the anthelmintic activity of this plant could be attributed to these compounds together or independently. Another previous chemical study on the plant has also resulted in the isolation of benzoquinones, anthraquinones, and a triterpenoid saponins as well as new antifungal dialkylbenzoquinone analog from the fruits of the plant [35]. The mechanism of action of active compounds of *Myrsine africana* is not understood. But probably these compounds have direct effect on the larvae.

Conclusion

The 80% Hydro alcoholic leaf extract of *Myrsine africana* possess anthelmintic activity that may support the usage of these plants by local traditional healers to treat hook worm infection. However, further research is required to study toxicity, mechanism of action, isolation identification and structure determination of pure chemical constituents and the effectiveness of the plant *in vivo* before it can be recommended for use in humans.

Conflict of interest

The authors declared no conflict of interest.

References

1. Koehler P (2001) The biochemical basis of anthelmintic action and resistance. *Int J Parasitol* 31: 336-45.
2. Patel J, Kumar GS, Deviprasad SP, Deepika S, Qureshi MS (2011) Phytochemical and anthelmintic evaluation of *Lantana camara* (L.) Var. *Aculeate* leaves against *pheretimaposthuma*. *J Global Trends Pharm Sci* 2: 11-20.
3. Partap S, Kumar S, Kumar A, Sharma NK, Jha KK (2012) In-vitro anthelmintic activity of *Luffacylindrica* leaves in Indian adult earthworm. *J Pharmacognosy and Phytochemistry* 1: 27-30.
4. Tandon V, Yadav AK, Roy B, Das B (2011) Phytochemicals as cure of worm infections in traditional medicine systems. *Emerging trends in zoology*, Narendra Publishing House, New Delhi 351-78.
5. Merid Y, Hegazy M, Mekete G, Teklemariam S (2001) Intestinal helminthic infection among children at Lake Awassa area, south Ethiopia. *Ethiop J Health Dev* 15: 31-7.
6. Kuma F, Birhanu T, Hirpa E (2015) Advanced Review on Anthelmintic Medicinal Plants. *Rep Opinion Children* 7: 6-16.
7. Deb Prashanta KR, Ghosh R, Das S, Bhakta T (2013) In-vitro Anthelmintic activity of *Acorus Calamus* leaves. *Asian J Pharmaceutclin Res* 6: 135-7.
8. Sissay M, Menbereleul M (2014) Prevalence of Opportunistic Intestinal Parasitic Infection among HIV/AIDS Patients Attending Othona Hospital, WolayitaSodo, Southern Ethiopia. (Doctoral dissertation) Harmaya: University of Ethiopia.
9. Mengistu A, Gebre-Selassie S, Kassa T (2007) Prevalence of intestinal parasitic infections among urban dwellers in southwest Ethiopia. *Ethiop J Health Dev* 21: 12-7.
10. BelaynehA, Asfaw Z, Demissew S, Bussa F (2012) Medicinal plants potential and use by pastoral and agro-pastoral communities in ErerValley of BabileWoreda, Eastern Ethiopia. *J Ethnobiol Ethnomed* 8: 12-6.
11. Dolinská M, Ivanišínová O, Königová A, Várady M (2014) Anthelmintic resistance in sheep gastrointestinal nematodes in Slovakia detected by in-vitro methods. *BMC veterinary research* 10: 233.
12. Eguale T, Giday M (2009) In vitro anthelmintic activity of three medicinal plants against *Haemonchus contortus*. *International Journal of Green Pharmacy (IJGP)* 3: 1-6.
13. Artho R, Schnyder M, Kohler L, Torgerson PR, Hertzberg H (2007) Avermectin-resistance in gastrointestinal nematodes of Boer goats and Dorper sheep in Switzerland. *Vet Parasitol* 144: 68-73.
14. Bizimenyera ES, Githiori JB, Eloff JN, Swan GE (2006) In vitro activity of *Peltophorum africanum* Sond. (Fabaceae) extracts on the egg hatching and larval development of the parasitic nematode *Trichostrongylus colubriformis*. *Veterinary Parasitology* 142: 336-43.
15. Alawa CBI, Adamu AM, Gefu JO, Ajanusi OJ, Abdu PA, et al. (2010) In vivo efficacy of *Vernonia amygdalina* (compositae) against natural helminth infection in Bunaji (*Bos indicus*) calves. *Pak Vet J* 30: 215-8.
16. Nelson G (1996) *The Shrubs and Woody Vines of Florida: A Reference and Field Guide*. Pineapple Press Inc, USA.
17. Anthony JP, Fyfe L, Smith H (2005) Plant active components—a resource for antiparasitic agents?. *Trends in parasitology*. 21: 462-8.
18. Laelago T, Yohannes T, Lemango F (2016) Prevalence of herbal medicine use and associated factors among pregnant women attending antenatal care at public health facilities in Hossana Town, Southern Ethiopia: facility based cross sectional study. *Archives of Public Health* 74:7.
19. Zerabruk S, Yirga G (2012) Traditional knowledge of medicinal plants in Gindeberet district, Western Ethiopia. *South African Journal of Botany* 78: 165-9.
20. Hedberg I, Edwards S, Demisew S (2003) *Flora of Ethiopia and Eritrea*. Addis Ababa, Ethiopia 4: 64-9.
21. Harborne JB (1973) *Phytochemical methods: A guide to modern techniques of plant analysis*. Chapman and Hall Ltd, London 279.
22. Bulbul L, Uddin J, MojumderSushanta S, Tanni S, FerdowshiNipa A, Baul S (2013) Phytochemical investigation and evaluation of antiemetic & anthelmintic activities of *Polygonum lapathifolium* roots extract. *Int J Pharm Life Sci* 4: 2632-7.
23. Mori H (1955) A new method for culturing hookworm. *YonagoActa Med* 1: 177-9.
24. Gill JH, Redwin JM, Van Wyk JA, Lacey E (1995) Avermectin inhibition of larval development in *Haemonchus contortus*—effects of ivermectin resistance. *International journal for parasitology* 25: 463-70.
25. Institute of Laboratory Animal Resources (ILAR) Committee on Care, Use of Laboratory Animals and National Institutes of Health (US) Division of Research Resources (2011) *Guide for the care and use of laboratory animals*. National Academies, Washington, DC, United States.
26. Al-Shaibani IRM, Phulan MS, Shiekh M (2009) Anthelmintic activity of *Fumariaparviflora* (Fumariaceae) against gastrointestinal nematodes of sheep. *Int J Agric Biol* 11: 431-36.
27. Githigia SM, Thamsborg SM, Maingi N, Munyua WK (2005) The epidemiology of gastrointestinal nematodes in Goats in the low potential areas of Thika District, Kenya. *Bulletin of Animal Health and Production in Africa* 53: 5-12.
28. Gachathi FN (1993) *Animal Diseases and Plants used*. English Kikuyu Botanical Dictionary. Amref Printing Department, Nairobi.
29. Beentje HJ (1994) *Kenya Trees, Shrubs and Lianas*. National Museums of Kenya, Nairobi.
30. Muthee JK, Gakuya DW, Mbaria JM, Kareru PG, Mulei CM et al. (2011) Ethnobotanical study of anthelmintic and other medicinal plants traditionally used in Loitokitok district of Kenya. *Journal of Ethnopharmacology*, 135: 15-21.
31. Desta B (1995) Ethiopian traditional herbal drugs. Part I: Studies on the Toxicity and Therapeutic activity of Local Taenicidal Medications. *J Ethnopharmacol* 45: 27-33.
32. Kakrani HK, Kalyani GA (1983) Experimental evaluation of anthelmintic and purgative activity of *Myrsine africana* fruits. *Anc Sci Life* 3: 82-4.
33. Suman A, Kumar DG, Kumar BD, Raj CR, Matushree VB (2011) Preliminary phytochemical investigation and anthelmintic activity of *Acacia suma* (Roxb) barks. *Int Res J Phar* 2: 136-41.
34. Dash S, Das C, Sahoo DC (2010) Phytochemical and anthelmintic screening of crude bark extract of *Adenanthera pavonina* Linn. *Int J Compr Pharm* 2: 1-4.
35. Manguro LOA, Midiwo JO, Kraus W (1997) Triterpenoids and steroids of *Myrsine africana* leaves. *Planta medica*. 63: 290.