

DNA Sequencing of Chilling Tolerant *Phaseolus Vulgaris L.* and Chilling *Sensitive Vigna Unguiculata L.*

Mallika R¹ and Muthuchelian K^{2*}

¹Department of Biochemistry, V.V.Vanniaperumal College for Women, India ²Department of Bioenergy, School of Energy and Environmental Sciences, Madurai Kamaraj University, India

*Corresponding author: Mallika R, Department of Biochemistry, V.V.Vanniaperumal College for Women, Virudhunagar-626001, India, Tel: 9486513038, Email: mallikaswathi01@gmail.com

Citation: Mallika R, Muthuchelian K (2019) DNA Sequencing of Chilling Tolerant *Phaseolus Vulgaris L*. and Chilling Sensitive *Vigna Unguiculata L*. J Advan Plant Sci 2: 102

Article history: Received: 22 January 2019, Accepted: 26 February 2019, Published: 28 February 2019

Abstract

The response of dehydrin gene in *P. vulgaris* L. and *V. unguiculata* L. to LNT (Low night temperature) stress was studied. It was inferred that dehydrin gene is induced to express the protein which confers an adaptation effect only in P.vulgaris L. but not in *V. unguiculata* L. LNT did not show any genetic variations when compared with a non LNT stress induced plant. Hence *P. vulgaris* L. is considered to be cold tolerant plant and *V. unguiculata* L. is a sensitive plant. *P. vulgaris* L. and *V. unguiculata* L. (LNT treated and control) can be analysed for other genes involved in cold stress. This observation hypothesizes that there is no significant gene level variations between LNT stress induced plant and control plants. It can also be hypothesized that variations may be occurring in its expression level to code for the protein responsible for making the plant tolerant to cold. Moreover the present study also suggested no significant gene level variations between *P. vulgaris* L. and *V. unguiculata* L. plants even though significant biochemical variations were observed when these plants were exposed to LNT stress.

Keywords: Dehydrin Gene; LNT Stress; Cold Tolerance; Gene Expression

Introduction

Crop plants are exposed to a range of external abiotic stress factors such as drought, salinity, cold, freezing, high light sensitivity, nutrient imbalances etc, Numerous cold induced genes have been isolated and characterized in a number plant species.

An increasing number of genes have been revealed to be induced during cold acclimatization [1,2]. Many of these genes encode proteins with known freezing tolerance. They encode newly discovered proteins such as the *Arabidopsis* COR6.6, COR15a and COR78 polypeptides or homologous of LEA proteins such as *Arabidopsis* COR47. The polypeptides encoded by these cold-responsive genes fall into a number of groups based on amino acid sequence similarities, but all share the property of being extremely hydrophilic. In addition, many have relatively simple amino acid compositions (i.e. are composed largely of a few amino acids) have repeated amino acid sequence motifs and remain soluble upon boiling in dilute aqueous buffer. Among the highly expressed cold responsive genes of *Arabidopsis* are the COR genes also designated LTI (low temperature induced), KIN (cold-inducible) and RD (responsive to desiccation). The COR genes comprise four gene families, each of which is composed of two genes that are physically linked in tandem array. The COR78, COR15 and COR6.6 gene pairs encode newly discovered polypeptides and the COR47 gene pair encodes homologous of LEA group II proteins (also known as dehydrins and LEA D11 proteins). Recent studies indicate that COR15a acts in concert with other COR genes to enhance freezing tolerance [3,4].

LEA proteins are expressed at different stages of late embryogenesis in seed embryos and under various conditions of stress including desiccation. LEA proteins are found in different tissues and in all cell types. They have been found to accumulate in cytoplasm and plastids. The very nature of this wide cellular distribution infers a protective function [3]. Furthermore, LEA proteins and heat shock proteins have been shown to be involved in protecting macromolecules such as enzymes and lipids. Dehydrins (LEA D11 family) are proteins that occur in plants due to dehydration, low temperature, and osmotic stress. Earlier Inheritance studies including QTL analysis in crop plants revealed apparent co-segregation of Dhn genes with phenotypes associated with drought and freezing [5,6]. Dhns are unified by the presence of one or more copies of a putative amphipathic α -helix forming domain (The K-segments) which is highly conserved in higher plants. This and other distinct domains of Dhns including a phosphorylatable (the S-segment) and an N- terminal consensus sequence (the Y-segment) are pieced together in a consistent manner, interspersed by other lesser conserved and usually repeated domains (the F-segments) [7,8].

The assembly of domains into numerous, yet consistent permutation has resulted in a range of Dhn polypeptide lengths from 82 to 575 amino acid residues. The number of occurrences of the K-segment varies from one to 11 within a single polypeptide. The bulk of the Dhn polypeptide in most cases contains regions or domains (the F-segments) that are rich in Gly and polar amino acids (especially Thr) and are tandemly repeated between K-segments. But there are contrary examples were the F-segments located between the K-segments are rich in other amino acids or do not exit as tandem repeats. For example, the F-segments located between K- segments in all SK3 Dhns are not Gly rich, but in many cases are rich in Pro and Ala. Because of this distinction and the fact that the SK3 and some other Dhns tend to contain high percentage of acidic residues, it has been proposed that the Dhn family may contain biochemically distinct acidic sub groups. Two studies have clarified the location of Dhns in the cytoplasm [10]. Most Dhns contain putative bipartite nuclear targeting signal sequences. The predicted molecular weights of Dhns based on amino acid sequences are invariably less than their apparent molecular weight SDS-PAGE. This anomaly is also observed with Dhns translated in-vitro, retarded migration thus seems to be due at least in part to secondary structure in 0.1% SDS. A ~35 kDa Dhn of V. unguiculata L. which is associated with an increment of chilling tolerance during seedling [10]. A simple interpretation of these observation is that Dhns are lipid binding proteins possibly, Dhns and other LEA and COR (cold responsive) proteins function in a lipoprotein environment, at an interface between phospholipids bilayers and aqueous compartment. Dehydrins are a D11 family of late embryogenesis abundant proteins that are induced in vegetative tissues in response to low temperature. The main aim is to isolate the dehydrin gene from cold stress induced in-vitro cultured P. vulgaris L. and V. unguiculata L. plants and to analyse if any sequence variations occurs in the gene level.

Materials and Methods

P. vulgaris L. (Sel.9) and *V. unguiculata* L. (P.152) seedlings were grown under controlled climatic condition (in a polyhouse) by irradiance of 1500 µmol m⁻²S⁻¹ (PAR), temperature of 28 °C and 70% RH for 7 days. The plants were watered regularly. The plants were classified into two groups according to the treatment. One group of plants was continuously grown under ambient irradiance and temperature controlled polyhouse, while the other group of plants were subjected to LNT (low night temperature) treatment of 15 °C by transferring the plants to chilling chamber and treatment was given for 12 h daily (each night from 18:00h to 06:00h) for 5 days. During daytime the plants were grown under polyhouse condition. On 13th day, total RNA was isolated.

RNA isolation and Amplification

100 mg of plant tissue was taken for the RNA isolation. The plant tissue was homogenized with 450 μ l of RLT buffer and 150 μ l of RLC buffer with the help of the homogenizer. The lysate was transferred into the QIA shredder spin column (lilac tube). The spin column was centrifuged at maximum speed for 3 min. After centrifugation, the supernatant of the flow through was transferred to the new microfuge tube without disturbing the cell debris. 225 μ l of 100% ethanol was added into the sample and it was mixed immediately by pipetting. Then the sample was applied into the mini spin column and centrifuged for 1 min. at 10,000 rpm. The flow through was discarded and 700 μ l of RW1 wash buffer was added to the column. The tube was centrifuged for 1 min. at 10,000 rpm. In the next step the flow through and the collection tube were discarded. The column was placed in a fresh collection tube and 500 μ l of RPE was added into the column and centrifuged for 1 min. at 10,000 rpm to wash the column. The column was centrifuged again for 1 min. to dry the membrane to avoid ethanol. The flow through and the collection tube and centrifuged for 2 min. at 10,000 rpm. The isolated RNA was visualized in the 1.2% formaldehyde agarose gel electrophoresis. The isolated RNA was amplified by RT-PCR. 12 μ l of amplified PCR product was loaded in the 1% agarose gelelectrophoresis [11,12].

Purification of PCR Products (Qiaquick PCR purification kit protocol)

5 volumes of PB buffer and 1 volume of PCR sample were taken in Qiaquick kit tube. Qiaquick spin column was placed in a provided 2ml collection tube. To bind DNA, the sample was applied to the Qiaquick column and centrifuged for 30-60 seconds. The flow through was discarded and the Qiaquick column back was placed into the same tube. 0.75 ml PE buffer was added to the Qiaquick column and centrifuged for 30-60 seconds. The Qiaquick column was placed in a clean 1.5 ml micro centrifuge tube and the DNA was eluted with 40 μ l elution buffer.

Cloning and Sequencing

The purified PCR products were cloned using QIAGEN PCR cloning plus kit as described by the manufacturer. Clones were selected and isolated plasmids with insert were sequenced with M13 Sequencing Primers using ABI biosystems automated sequencer (Macrogen Genomics, Korea).

Database Searching

Nucleotide database was searched with the sequences obtained using NCBI BLAST (Blastn) tool [13].

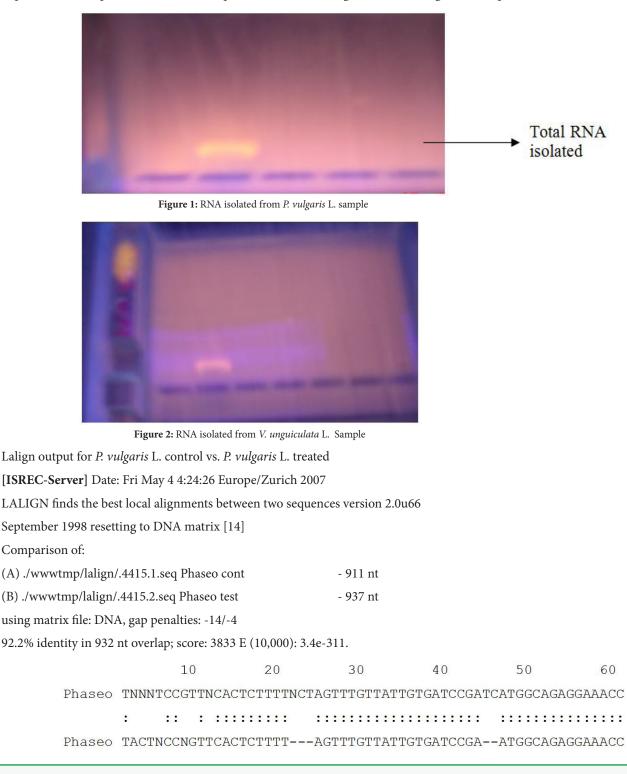
Sequence Analysis

Sequences of both the control and treated *Phaseolus* and *Vigna* species were analyzed using pairwise sequence alignment LALIGN software from EMBNET server [14].

Results

The total RNA isolated from *P. vulgaris* L. and *V. unguiculata* L. (cold stress treated and control) plants produced a distinct band and it was observed in 1.2% FAGE on UV illumination (Figure 1 and 2). Discrete band for *P. vulgaris* L. control dehydrin had the size of 911 bp, while treated dehydrin had 937 bp. In *V. unguiculata* L. control dehydrin (Dhn1) mRNA had 908 bp, while treated dehydrin (Dhn1) mRNA had 982 bp, in 1% agarose gel on UV illumination after loading the RT-PCR product amplified using Dhn primers.

The amplified products were purified and sequenced. The sequences were obtained from automated sequencer. The cold stress induced dehydrin gene of *P. vulgaris* L. and *V. unguiculata* L. and database sequence of control *P. vulgaris* L. and *V. unguiculata* L. plants were genetically analysed using Bioinformatics tools. Pair wise alignment indicated 98% similarity between the cold stresses induced dehydrin gene of *P. vulgaris* L. and *V. unguiculata* L. and database sequence of control *P. vulgaris* L. and *V. unguiculata* L. This genetic analysis concluded that there was no genetic variation between the LNT stress induced *P. vulgaris* L. and *V. unguiculata* L. plant when compared with database sequence of control *P. vulgaris* L. and *V. unguiculata* L.



20	30	4	0 5	50	60 70
	70 8				
Phaseo CAGAACA					
Phaseo CAGAACA					
FIIaSeo CAGAACA	80				120
	00		100	110	120
Phaseo TTTCTC	130 14 GGTAAGGGAAA		150 GAAGAAGAG-	160 AAGCCAGO	
Phaseo TTTCTC	GGTAAGGGAAG	CCGCTAGTAG	GAAGAAGAG	CCGGAAGCCAG	GAAGAGGTGATCG
130	140	150	160	170	180
180	190	200	210	220	230
Phaseo TCACCG	AGTTTGAAAAG	ATCACAGTGT	CAGAGGAGAG	GAAGGAAGAGGA	AAGAGAAGAA
:::::			•••••		
Phaseo TCACCG	AGTTTGAAAAG	ATCACAGTGT	CAGAGGAGAG	GAAGGAAGAGGA	AGAGAAGCACAG
190	200	210	220	230	240
240) 250	260	270	0 280	290
Phaseo GCACAGO	CTTTTAGAAAA	AGCTTCACGA	TCTGACAAG	CCGGTTATCCC	IGATAGCTCTTCA
Phaseo GGAACTC					
250	260	270	280	290	300
300)	310	320	330	340
Phaseo AGCGAAG	GGAAGGI	AGAAGACGGG	GAGAAAAAGA	AAGAAGAAGAA	GAAGGAGAAGAGG
::::::					
Phaseo AGCGAAG 310	GGATTCAGAGGA 320	AGAAGACGGGG 330	GAGAAAAAGA 340	AAGAAGAAGAA 350	GAAGGAGAAGAGG 360
350	360	370	380	390	400
Phaseo AGAAGAA					
Phaseo AGAAGAA	AGATACGA	AGAGAAAATA	GAGGGTTAT	CACAAGGAAGA	CACGAGTGTTCCA
370	380) 39	0 4	00 41	0 420
410	420	120	110	450	160
410 Phaseo GTGGAGA					
					GGAGAAGAIIAAG
Phaseo GTGGAGA					
	30 440				70 480

	470	480	490	500	51	10 5	520
Phaseo	GAGAAGCTA	CCAGGGCA	CAAGAATO	GAGGAGGCG	GCAGCTCC	FCCTCCGCCA	ACCACCTGCT
Phaseo	GAGAAGCTA 490		CAAGAATC 00	GAGGAGGCG 510	GCAGCTCC 520	ICCTCCGCCA 530	ACCACCTGCT 540
	530	540	550	560	57	70 5	80
Phaseo	GCCACCTCA	TTGATCGG	AACATGAA	GGAGGCGA	AGGAGAAGA	AAGTATATI	GGAGAAGAT
Phaseo	GCCACCTCA	TTGATCGG	AACATGAA	GGAGGCGA	AGGAGAAGA	AA-TATATI	GGAGAAGAT
	550	5	60	570	580	590	600
	590	600	61	.0	620	630	640
Phaseo	AAAAGA ::::::	GAAGCTTC	CTGGTTAC	CACTCAAA	GACAGAGGA	AGGACGTAGA	AAGAAAAGC
Phaseo	TATAAAAGA	GAAGCTTC	CTGGTTA	CCACTCAAA	GACAGAGG	AGGACGTAG	AAGAAAAGC
	61	.0	620	630	640	650	660
	650		660	670	680	690	700
Phaseo	GGGAGATG-	GGTCA	CTGAGAT	GAGAGAAG	GGTTGTGT	rgtggggttt	TGGTTGGATG
Phaseo	GGGAGATGC	CTAGGGTCA	CTGAGAT	GAGAGAAG	GGTTGTGT	IGTGGGGTTI	TGGTTGGATG
	67	0	680	690	700	710	720
	71		720	730	740	750	760
Phaseo						TGTGGGCTT	TCTTATCTTT
Dhagaa							тсттатсттт
Pliaseo		30		750			70
	1.	30	740	150	76	0 7	70
	7	70	780	790	8	00	810
Phaseo	TGTGCTTCI	TCTTCAT	GCATACTT	FACTTTTTA	GCGGA	TTATTATTA	ITGTGTATGT
Phaseo	TGTGCTTCI	TCTTCATO	GCATACTT	TACTTTTTA	GCGGCCAA	TTATTATT-	ITGTGTATGT
j	780	790	800	810	820	0 8	330
	820	830	840	850)	860	870
Phaseo	ATCATCTCI	TGCATATA	CAAAAAG	CTTTCGTAC	CTT	TCCTC-TTT	CTTTTTCTCA
					: ::		: ::::::
Phaseo	ATCATCTCI	TGCATATA	CAAAAAG	CTTTCGTAC	CAGGTCTT	TCCTCNTTT	CNNTTTCTCA
	840	850	860	870	81	80 8	390

Phaseo TAACTTGTTTGATGT---AATGTATGGANANA Phaseo TAACTTGTTTGATGTNANAATGTATGGAAAGA 54.7% identity in 802 nt overlap; score: 383 E(10,000): 1.6e-23 Phaseo GAGAGCAGTGAGGTG-GAGGTCCAGGATCGTGGAGTTTTGACTTTCT-CGGTAAGGGAAA : : Phaseo GAGCGCAGAGAGCAGTGAGGT---GGGTCCAGGATCGTGGAGTTTTGACTTTCTCGGTAA Phaseo ---AAGCCG-TAGGAAGAAGAAGAAGCCAGGAAG--AGGTGATCGTCACCGAGTTTGAA--.. . .. :: Phaseo GGGAAGCCGCTAGTAGGAAGAAGAGCC-GGAAGCCAGGAAGAGGTGATCGTCACCGAGTT Phaseo -- AAGATCACAGTGTCAGAGGAGAGAGAAGGAAGAGAAGAAGAAGAAGCACAGCCTTTTAGA :: Phaseo AAAGCTTCACGATCTG----ACAAGCCGGTTATCCC-TGATAGCTCTTCAAGCGAAG-GA ::: :: 111 11 : Phaseo ACTCCTTTTAGAAAAGCTTCACGATCTGACAAGCCGGTTATCCCTGAGCTCTTCAAGCGA :::: Phaseo AAGATTACGAGAGAAAATAGAGGGTTATCACA-AGGAAGACACGAGTGTTCCAGTGGAGA : : Phaseo AAGATA---CGAGAGAAAATAGAGGGTTATCACAAGGA-AGACACGAGT----GTTCCAG

J Adv Plant Sci

 380
 390
 400
 410
 420

Phaseo GCTACCAGGGCACAAGAATGAGGAGGCGGCAGCTCCTCCTCCGCCACCACCTGCTGCCAC : : Phaseo GAAGCTACCAGGGCACAAGAATGAGGAGGCGGCAGCTCCTCCTCCGCCACCACCTGCTGC

Phaseo AGATGGGTCACTGAGATTGAGAGA-AGGGTTGTGTGTGGGGGTTTGGTT-GGATGATTGT ::

Phaseo CTTCTTCTTCATGCATACTTTACTTT--TTAGCGGATT---ATTATTATTGTGTATGTAT Phaseo -TGCTTCTTC-TTCATGCAT-ACTTTACTTTTTAGCGGCCAATTATTATTTTGTGTATGT

Phaseo CATC-TCTTGCATATACAAAAA ::: ::: ::: ::: ::: Phaseo -ATCATCTCTTGCATATACAAA 840 850 55.4% identity in 578 nt overlap; score: 258 E(10,000): 4.3e-13

Phaseo AGAAGA-AGAAGATTACGAGAGAAAATAGAGGGTTATCACAAGGAAGACACGAGTGTTCC ::: :: : :::: Phaseo ATACGAGAGAAAAT--AGAGGGTTATCACAAGGAAGACACGAGTG---TTCCAGTGGAGA Phaseo AGTGGAGAAAGTG-AGGTTGTGGAAGGGAAAAGAA----GGGGGGCCATTCCTGGAGAAGA : : ::: ::: : : ::::: Phaseo AAGTGAGGTTGTGGAAGGGAAAGAAGGGCCGACATTCCTGGAGAAGATTAA-GGAGAAGC Phaseo TTAAGGAGAAGCTACCAGGGCACAAGAATGAGGAGGCGGCAGCTCCTCCTCCGCCACCAC 1 1 11 1 11 11 Phaseo T--ACCAG--GGCACAAG----AATGAGGAGGCGGCAGCTCCTCCTCCGCCACCACCTG Phaseo CTGCTGCCACCTCATT--GATCG----GAACATGAAGGAGGCGAAGGAGAAGAAAGTATA Phaseo CTGCCACCTCATTGATCGGAACATGAAGGAGGCGAAGGAGAAGAATATATTGGAGAAGA Phaseo TTGGAGAAGATAAAAGAGAAGCTTCCTGGTTACCACTCAAAGACAGAGGAGGACGTA-GA 1 11 1 1 11 Phaseo TTATAAAAGAGAAGCTTCCTGGTTACCACT---CAAAGACAGAG-GAGGACGTAGAAAGA Phaseo AA-----GAAAAGC--GGGAGA-TGGGTCACTGAGATTGAGAGAAGGGTTGTGTGTGGG :: :: :

Phaseo AAAGCGGGAGATGCTAGGGTCACTGAG--ATTGAGAGA-AGGGTTGTGTGTGTGGGGGTTTG

Volume 2 | Issue 1

690 700 710 720 730 740 :: :::: : :: : Phaseo GTT-GGATGATTGTGGTGTGCTTTGTTTTCATTCATCATATCAAACTTG----TGGGC 720 730 740 750 760 750 760 770 780 790 800 : : : : : : ::: Phaseo TTTCTTATCTTTGTGCTTCTTCTTCATGCATACTTTACTTT--TTAGCGGCCAATTATT 770 780 790 800 810 820 810 820 830 Phaseo ATTATTATTGTGTATGTATCATCTCTTGCATATACAAA : ::::: Phaseo ATTTTGTGTATGTATC-ATC-TCTTGCATATACAAAAA 830 840 850 860 Lalign output for V. unguiculata L. control vs. V. unguiculata L. treated [ISREC-Server] Date: Fri May 4 4:14:27 Europe/Zurich 2007 LALIGN finds the best local alignments between two sequences version 2.0u66 September 1998 resetting to DNA matrix [14] 381 resetting to DNA matrix Comparison of: (A) ./wwwtmp/lalign/.4659.1.seq Vigna cont - 908 nt (B) ./wwwtmp/lalign/.4659.2.seq Vigna test - 982 nt using matrix file: DNA, gap penalties: -14/-4 96.0% identity in 916 nt overlap; score: 4137 E(10,000): 0 70 20 30 40 50 60 GAAACTGATG--AATATGGCAACCCGGTTC-ATGCAGCAAGTGTCCTCGGGCCTCCACCA Vigna Vigna GAAACTGATGNNAATATGGCAACCCGGTTCNATGCAGCAAGTGTCCTCGGGCCTCCACCA 20 30 40 50 60 70 80 90 100 110 120 130 Vigna CCGGTGGTCTTGGCCTGGATGACACTAACAAGCAACATGATACCAGTAATGTCTACGGTG CCGGTGGTCTTGGCCTGGATGACACTAACAAGCAACATGATACCAGTAATGTCTACGGTG Vigna 80 90 100 110 120 130

660

670

680

690

700

710

	140)	150	160	170	180	190
Vigna				TGGCATAGG		AGACAGCACGO	
2							
Vigna	CAGACACO	CCGTAGAC	CACGGAACTT	TGGCATAGG	IGACACCGGT	AGACAGCACGO	GAACTAC
	14	10	150	160	170	180	190
	200)	210	220	230	240	250
Vigna	CGGTGGT	TTACTGO	GTGACACCGG	TAGACAATAT	IGGCACTACC	GGAGGCTTTAC	CCGGTGA
				••••			
Vigna	CGGTGGT	TTACTGO	TGACACCGG	TAGACAATAT	IGGCACTACC	GGAGGCTTTAC	CCGGTGA
	20	0	210	220	230	240	250
	260)	270	280	290	300	
Vigna	CACCGGGA	AGACAACA	ATGGGACTAC	CGGTGGTTT	FACCGGTGAC	ACTGGG	FAACAAC
						:::::	
Vigna	CACCGGGA	AGACAACA	ATGGGACTAC	CGGTGGTTT	FACCGGTGAC	CTTCACTGGG	FAACAA C
	26	50	270	280	290	300	310
	310	320	330	340	350	360	
Vigna	ATGGGACT	TACCGGT	GTTTTACGT	GACACTGGG	AGACAACATG	GGACTACTGG	FGGTT TT
Vigna			GGTTTTACGI			GGACTACTGG	IGGTTTT
	3	20	330	340	350	360	370
	370	380	390	400	410	420	
Vigna						GGAGACAACAT	GCGACT
, Lyna							
Vigna						GGAGACAACAT	
<u>-</u>		30			410		
	430		440	450	460	470	
Vigna	ACCGGC					GGGAGGCCCTT	ACGTTG
Vigna	ACCGGCC	TCCGAAA	GCGGCTTTA	CTGGTGGTGA	CACTGGTCT	GGGAGGCCCTT	ACGTTG
1 (Kal)				460		272722	490
	480					530	
Vigna	GAGCCAA	CACCGCAC	CACAGGGACT	GGTCCTAGA	AGTGGCACAG	GTGGCAGCGC	CTATGGA
100/0101/02 00000						GTGGCAGCGC	

J	Adv	Plan	nt Sci
---	-----	------	--------

		500	510	520	530	540
	540	550	560	570	580	590
Vigna						CACGGGGGGGAGC
VIGIIa						
Vigna						CACGGGGGGGAGC
-	550	560	570	580		
	550	560	570	200	590	600
	600	610	620	630	640	650
Vigna	ACACTCAC	GTGATGAAAGG	GTATGGAAGGG	AGTATCGTGA	GCAATATGGG	TAGTCTCGTGGA
Vigna	ACACTCAG	GTGATGAAAGG	GTATGGAAGGG	AGTATCGTGA	GCAATATGGG	TAGTCTCGTGGA
	610	620	630	640	650	660
	660	670	680	690	700	710
Vigna	AAGATCAT	GACAAGAAAG	GGATAGTGGA	CAAGATTAAG	GAGAACCCGG.	AGGACACAGTGA
	670	680	690	700	710	720
	720	730	740	750	760	770
Vigna	CAANCAAG	TACATCATGO	GTGTGCATGC	CATGCGTATAT	ATACGGGTAG	TATAATTAAAG-
	::: ::::					
Vigna	CAA-CAAG	GTACATCATGO	GTGTGCATGC	ATGCGTATAT	ATACGGGTAG	TATAATTAAAGG
	730	740	750	760	770	780
		780	790	800	810	820
Vigna		ATGTTATA	ATTGTTGTGTI	TTTGAATAAG	TTTGCTGCAT	ATATACGTACTC
		:::::::				
Vigna	GGGGTTTA	ACTATGTTATA	ATTGTTGTGTGTI	TTTGAATAAG	TTTGCTGCAT	ATATACGTACTC
	790	800	810	820	830	840
	830	840	850	860	870	880
Vigna	GTACACT	GTCGTTCTCG	TGTAGGNTAT	GTGGTGGATCI	TGTATATNGG	TTNATAGTAAAG
Vigna	GTACACT	GTCGTTCTCG	TGTAGG-TAT(GTGGTGGATCI	TGTATAT-GG	TT-ATAGTAAAG
	850	860	870	880	890	900
	890	900				
Vigna		AATTGCATC				

Vigna ANGGAATAATTGCATG

: :::::: Vigna A-GGAATAATTGCATG 910

66.1% identity in 599 nt overlap; score: 868 E(10,000): 6.1e-64

	170	180	190	200	210	220	
Vigna	GACACCGGT	AGACAGCAC	GGAACTACO	GGTGGTTTI	TACTGGTGAC	ACCGGTAGA	CAATAT
				::::			:: :
Vigna	GACACCCGT	AGACAC	GGAACT	TTGGCAI	TAGGTGAC	ACCGGTAGA	CAGCAC
	140	1	50	160	1	70	180
	230	240	250	260	270	280	
Vigna	GGCACTACC	GGAGGCTTT	ACCGGTGAC	CACCGGGAGA	ACAACATGGO	;ACTACCGGT	GGTTTT
							:: :::
Vigna	GGAACTACCO	GGTGGTTTT	ACTGGTGAC	CACCGGTAGA	ACAATATGGC	ACTACCGGA	GGCTTT
	190	200	210) 22	20 2	230	240
		200	010	200	222		240
Mi an a	290				330	CIIICAC	340
Vigna	ACCGGTGACA						
Vigna	ACCGGTGAC						
vigila	250		270				00
	250	200	210	20	0 2.	, o 5	
	350) 3	60	370	380	390	400
Vigna	GAGACAACA					ACATGGCCG	ACTAC
Vigna	GTAACAACA	GGGACTAC	CGGTGGTTT	TACGTGACA	CTGGGAGACA	AACATGGG	ACTAC
	310	320	330	34	0 3!	50	360
		4	10	420	430	440	
Vigna	CGGTG	ACA	CTGGGAGAC	CAACATGC	CGACTACCGG	CGGCTTT	ACT
							::
Vigna	TGGTGGTTT						
	370	38	0 3	390	400	410	420
	450		1.00	470	100	100	
T 7 '	450		460	470	480	490	
Vigna	GGTGGTGAC						
17: and							
Vigna	ACATGCGAC				460	ACACTGGTC 470	TGGGHG
	430	44	4	50	400	470	

	500	510	520	530	5	40
Vigna	GGACTGG	[CCTAGAAGT	GG-CACAGG	IGGCAGCGCC'	raTggat	CGGGTGGTTA
		: : : :	: :::::	: : : ::		: :::::
Vigna	GCCCTTACGT	[GCCAACACC	GCACACAGG	-GACTGGTCC	TAGAAGTGGCA	CAGGTGGCAG
4	80 490	500	0	510	520 5	30
	550 50	60	70	580	590 6	00
Vigna	TGGATGTGGA	ATCAGCTGGA	GCTGGGTAT	GGTATGAACA	CGGGGGGGAGCA	CACTCAGTGA
	: : ::::				: :::::	: : :
Vigna	CGCCTATGGA	-TCGGGTGGT	TATGGATGT	GGAATCA	GCTGGAGCT	GGGTATGGTA
	540	550	560	570	580	590
	61.0				65.0	6.60
	610 62			540		660
Vigna	TGAAAGGTATG	GAAGGGAGTA	\TCGTGAGC <i>I</i>	AT-ATGGGTA	AGTCTCGTGGA	AAG-ATCATG
			: :::			
Vigna	TGAACACG					
	600	61	LO e	520	630	640
	670	680	690	700	710	720
Vigna	A-CAAGAAAGG	GATAGTO	GACAAGATT	TAAGGAGAAC	CCGGAGGACAC	AGTG-ACAA
	: ::: : ::			:: ::	::::	
Vigna	AGCAATATGGG					
	650	660	670	680	690	
61.3% identity in 802	nt overlap; score:	865 E(10,000):	1.1e-63			
	140	150		160	170	180
Vigna	G3 G3 GGGGB3					TOO
	GACACCCGTA	GACACGG	AACTT	TGGCATA	GGTGACACCGG	
	GACACCCGTA			TGGCATA	GGTGACACCGG	TAGACAGCAC
Vigna						TAGACAGCAC
Vigna						TAGACAGCAC
Vigna	GACACCGGTA	GACAGCACGG	:::: AACTACCGG	::: : TGGTTTTACT	GGTGACACCGG	TAGACAGCAC :::::: : TAGACAATAT
Vigna	GACACCGGTA	GACAGCACGG	:::: AACTACCGG	::: : TGGTTTTACT	GGTGACACCGG	TAGACAGCAC :::::: : TAGACAATAT
Vigna Vigna	:::::: ::: GACACCGGTA 170 190	GACAGCACGG 180 200	:::: AACTACCGG 190 210	::: : TGGTTTTACT 200 220	GGTGACACCGG 210	GTAGACAGCAC :::::::::::::::::::::::::::::::::::
	GACACCGGTA 170 190 GGAACTACCG	GACAGCACGG 180 200 GTGGTTTTAC	XACTACCGG 190 210 TGGTGACAC	IGGTTTTACT 200 220 CGGTAGACAA	GGTGACACCGG 210 230	TAGACAGCAC TAGACAATAT 220 240 CCGGAGGCTTT
	:::::::: GACACCGGTA 170 190 GGAACTACCG :: :::::::	::::: ::: GACAGCACGG 180 200 GTGGTTTTAC : :: :::::	AACTACCGG 190 210 TGGTGACAC	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	GGTGACACCGG 210 230 TATGGCACTAC	TAGACAGCAC TAGACAATAT 220 240 CCGGAGGCTTT T T T T T T T T T T T T T T T T T T
Vigna	:::::::: GACACCGGTA 170 190 GGAACTACCG :: :::::::	::::: ::: GACAGCACGG 180 200 GTGGTTTTAC : :: :::::	AACTACCGG 190 210 TGGTGACAC	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	GGTGACACCGG 210 230 TATGGCACTAC	TAGACAGCAC TAGACAATAT 220 240 CCGGAGGCTTT T T T T T T T T T T T T T T T T T T
Vigna	GACACCGGTA 170 190 GGAACTACCG GGCACTACCG	GACAGCACGG 180 200 GTGGTTTTAC : :: ::::: GAGGCTTTAC	AACTACCGG 190 210 TGGTGACAC :::::::::	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	GGTGACACCGG 210 230 TATGGCACTAC :	TAGACAGCAC TAGACAATAT 220 240 CGGGAGGCTTT CGGTGGTTTT
Vigna	GACACCGGTA 170 190 GGAACTACCG GGCACTACCG	GACAGCACGG 180 200 GTGGTTTTAC : :: ::::: GAGGCTTTAC	XACTACCGG 190 210 TGGTGACAC CCGGTGACAC 250	IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII	GGTGACACCGG 210 230 TATGGCACTAC ::::::::::: CATGGGACTAC 270	TAGACAGCAC TAGACAATAT 220 240 CGGGAGGCTTT TTT CCGGTGGTTTT 280
Vigna Vigna	GACACCGGTA 170 190 GGAACTACCG :: :::::: GGCACTACCG 230	::::: :::: GACAGCACGG 180 200 GTGGTTTTAC : :: ::::: GAGGCTTTAC 240 260	XACTACCGG 190 210 TGGTGACAC CCGGTGACAC 250 27(<pre>:::: : TGGTTTTACT 200 220 CGGTAGACAA ::: ::::::: CGGGAGACAA 260 28</pre>	GGTGACACCGG 210 230 TATGGCACTAC ::::::::::: CATGGGACTAC 270 0 290	TAGACAGCAC TAGACAATAT 220 240 CCGGAGGCTTT TIN TIN CCGGTGGTGTTTT 280 300
Vigna Vigna	:::::::: GACACCGGTA 170 190 GGAACTACCG :::::::::: GGCACTACCG 230 250	::::: :::: GACAGCACGG 180 200 GTGGTTTTAC : :: ::::: GAGGCTTTAC 240 260	XACTACCGG 190 210 TGGTGACAC CGGTGACAC 250 270 GACAACATGO	<pre>:::: : TGGTTTTACT 200 220 CGGTAGACAA ::: ::::::: CGGGAGACAA 260 28</pre>	GGTGACACCGG 210 230 TATGGCACTAC :::::::::: CATGGGACTAC 270 0 290 IGGTTTTACCG	TAGACAGCAC TAGACAATAT 220 240 CCGGAGGCTTT TIN TIN CCGGTGGTGTTTT 280 300

13

	290	300	310	320	330	340
) 340		
Vigna				CGTGACACTGGG		
wast. on						
Vigna				GTGACACTGGG		
	350	360	370	380	390	400
	360 3	370 3	80	390 40	0 410)
Vigna				CATGGCCGACTA		
<u> </u>	:					
Vigna	CG			CATGCGACTA		
		410		430		40
	420	430	440	450	460	470
Vigna	ACATGCGACT	ACCGGCGG	CTTTACTGG	I GGTGACAC	CTGGTCTGGG	GAGGCCCTT
	:	:	:::::		: ::::	: ::
Vigna	GCGGCT	TTACTGGTGG	TGACACTGG	CTGGGAGGCCC	CTTACGT-TGCC	CAACACCGC
	450	460	470	480	490	500
	360 3	370 3	80 3	390 40	0 410	
Vigna	TGGTGGTTTT	AGGTGACACT	GGGAGACAA	CATGGCCGACTA	CCGGTGACACT	GGGAGACA
	:		••••			
Vigna	CG			CATGCGACTA		
		410	420	430	4	40
	100	120	4.4.0	450	100	470
Vigna	420	430	440	450 GGGTGACAC	460	470
Vigila	ACAIGCGACI					AGGCCCII
Vigna				CTGGGAGGCCC		
Vigila	450	460	470	480	490	500
			2.6.2		5.5.5	
	480		490	500	510	520
Vigna	ACGTTGGAGC	cCAACA	CCGCACACAC	G-GACTGGTCC	TAGAAGTGGCA	CAGGTGGC
		:::	:::::			: :::::
Vigna	ACACAGGGAC	TGGTCCTAGA	AGTGGCACAG	GTGGCAGCGCC	TATGGAT	CGGGTGGT
	510	520	530	540	55	0
	530	540	550	560	570	580
Vigna	AGCGCCTATG	GA-TCGGGTG	GTTATGGATC	TGGAATCAGCT	GGAGCTGGGTA	TGGTATGA
	: : ::					:: :
Vigna	TATGGATGTG	GAATCAGCTG	GAGCTGGGTA	TGGTAT	-GAACACGG	-GGGGAGC
	560	570	580	590	600	

14

	710	720	730	740	750
Vigna	GGAGGACACAG	TGACAANCAAG-	TACAT-CAT	GGGTGTGCATGC	ATGC-GTATATAT
	: : :::	::: :: :	: ::: :::		: ::::: :
Vigna	AGTGACAACAA	GTACA-TCATGG	GTGTGCATGCAT	GCGTATATATAC	GGGTAGTATAATT
	730	740	750	760 77	0 780
	760	770 78	0 790	800	810
Vigna	AC-GGGTAGTA	TAATTAAAGATG	TTATATTGTTGT	GTTTTTGAATAA	GTTTGC-TGCA
	: ::: ::				
Vigna	AAAGGGGGGTT	TACTATGTTA	TATTGTTGT-GT	-TTTTGAATAAG	TTTGCTGCATATA
	790	800	810	820	830
	710	720	730	740	750
Vigna	GGAGGACACAG	TGACAANCAAG-	TACAT-CAT	GGGTGTGCATGC	ATGC-GTATATAT
	:::::				: ::::: :
Vigna	AGTGACAACAA	GTACA-TCATGG	GTGTGCATGCAT	GCGTATATATAC	GGGTAGTATAATT
	730	740	750	760 77	0 780
		770 780		800	810
Vigna	AC-GGGTAGTA	TAATTAAAGATG	TTATATTGTTGT(GTTTTTGAATAA	GTTTGC-TGCA
Ser.					
Vigna					TTTGCTGCATATA
	790	800	810	820	830
	820	830	840	850 8	60 870
Vigna					GGATCTTGTATAT
Vigila	:: ::: ::				: :: : :: :::
Vigna					IGTATATGGT-TAT
<u> </u>		50 860			
	880	890			
Vigna	NGGTTNATAG	TAAAGANGGAAT			
Vigna	AG-TAAAGAG	GAATAATTGCAT			
	900	910			

Discussion

A growing number of genes have been shown to be induced during cold acclimatization. The cold stress induced dehydrin gene of *P. vulgaris* L. and *V. unguiculata* L. and database sequence of control *P. vulgaris* L. and *V. unguiculata* L. plants were genetically analysed using Bioinformatics tools. Pair wise alignment indicated 98% similarity between the cold stress induced dehydrin gene of *P. vulgaris* L. and *V. unguiculata* L. and database sequence of control *P. vulgaris* L. and *V. unguiculata* L. Dehydrin gene enhances tolerance to freezing stress in *Arabidopsis* and in Transgenic Arabidopsis plants over expressing multiple dehydrins were generated [7,15]. These findings showed that over expression of dehydrin results in increased freezing tolerance. This genetic analysis concluded that there was no genetic variation between the LNT stress induced *P. vulgaris* L. and *V. unguiculata* L. plant when compared with database sequence of control *P. vulgaris* L. and *V. unguiculata* L. plant

Volume 2 | Issue 1

This observation hypothesizes that there is no significant gene level variations between LNT stresses induced plants and control plants. It can also be hypothesized that variations may be occurring in its expression level to code for the protein responsible for making the plant tolerant to cold. Dehydrin gene produces the protein in response to LNT stress [16]. Moreover the present study also suggested no significant gene level variations between *P. vulgaris* L. and *V. unguiculata* L. plants even though significant biochemical variations were observed when these plants were exposed to LNT stress.

Conclusion

In this study, we have attempted to study the response of dehydrin gene in *P. vulgaris* L. and *V. unguiculata* L. to LNT stress. From this study, it can be inferred that dehydrin gene is induced to express the protein which confers an adaptation effect only in *P. vulgaris* L. but not in *V. unguiculata* L. LNT does not show any genetic variations when compared with a non LNT stress induced plant. Hence *P. vulgaris* L. is considered to be cold tolerant plant and *V. unguiculata* L. is a sensitive plant. *P. vulgaris* L. and *V. unguiculata* L. (LNT treated and control) can be analysed for other genes involved in cold stress. The Dhn gene can be cloned and expressed. The expressed protein characteristics can also be studied. The Dhn gene can be genetically transferred to plants using gene transformation methods to produce stress tolerant plants. This will be very useful for agriculturists to increase crop productivity which will be a great boon for a country like India with agriculture as prime occupation.

References

1. Hughes MA, Dunn MA (1996) The Molecular Biology of Plant acclimation to low temperature. Exper Bot 47: 291-305.

2. Thomashow MF (1999) Plant cold acclimation: Freezing tolerance genes and regulatory mechanisms. Plant Physiol 50: 571-99.

3. Shantersign Clauclia C, Robert Tyler C, Lee S, Markley L (2005) Solution structure of a late embryogenesis abundant protein (LEA14) from Arabidopsis thaliana, a cellular stress related protein. Protien science 14: 2601-9.

4. Danyluk J, Perron A, Houde M, Limin A, Fowlor B, et al. (1998) Accumulation of acidic dehydrin in the vicinity of the plasma membrane during cold acclimation of wheat. Plant Cell 10: 623-38.

5. Borovskii, Stupnikova GB, Antipina AA, Douin CA, Voinikov VK, et al. (2000) Accumulation of dehydrin like proteins in the mitochondria of cold treated plants. Plant Physiol 156: 797-800.

6. Fowler DB, Limin AE, Ritchie JT (1999) Low temperature tolerance in cereals: model and genetic interpretation. Crop Sci 39: 626-33.

7. Allagulova CR, Gimalov FR, Shakirova FM, Vakhitov VA (2003) The plant dehydrins; Structure and putative functions. Biochem 68: 945-51.

8. Hinniger C, Caillet V, Michoux F, Tankslay S, Mccarthy J, et al. (2005) Isolation and Characterization of cDNA encoding three Dehydrins, expressed during Coffea Canephora Grain Development. Anna Bot 97: 755-65.

9. Abdelbagi Ismail M, Hall E, Close J (1999) Allelic variation of a dehydrin gene cosegregrates with LNT tolerance during seedling emergence. Plant Sci 96: 13566-70.
 10. Abdelbagi Ismail M, Hall E, Close J (1999) Purification and Partial characterization of a dehydrin involved in LNT Tolerance during seedling Emergence of cowpea. Plant Physiol 120: 237-44.

11. Brawerman G (1974) Methods in Enzymol 30 (Eds Moldave and Grossman, L) Academic Press New York 605.

12. Sambrook J, Fritsch EF, Maniatis T (1989) Molecular Cloning: a Laboratory Manual. Cold Spring Harbor Laboratory press.

13. Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, et al. (1997) Gapped BLAST and PSI-BLAST: a newgeneration of protein database search programs. Nucleic Acids Res 25: 3389-402.

14. Huang X, Miller W (1991) Advances in Applied Mathematics. Adv Appl Math 12: 373-81.

15. Lee Sc, Lee MY, Kim SJ, Jun SH, Kim SR (2006) Characterization of an abiotic stress-inducible dehdrin gene, OS Dhn 1, in rice (Oryza stiva L.). Molecules Cells 19: 212-8.

16. Wang W, Vinocur B, Altman A (2003) Plant responses to drought, salinity and extreme temperatures: towards genetic engineering for stress tolerance. Planta 218: 1-14.